

A linkage disequilibrium study of bipolar disorder and microsatellite markers on 22q13

Sherri G. Liang^a, A.Dessa Sadovnick^b, Ronald A. Remick^c, Paul E. Keck^d, Susan L. McElroy^d and John R. Kelsoe^a

^aDepartment of Psychiatry, University of California, San Diego and San Diego VA Healthcare System, La Jolla, California, USA; ^bDepartment of Medical Genetics, University of British Columbia, Vancouver, Canada; ^cDepartment of Psychiatry, St Paul's Hospital, Vancouver, Canada; ^dDepartment of Psychiatry, University of Cincinnati, Cincinnati, Ohio, USA

Correspondence to John R. Kelsoe, Department of Psychiatry 0603, University of California, San Diego, La Jolla, CA 92093, USA. E-mail: jkelsoe@ucsd.edu

Received 31 August 2001; accepted 22 January 2002

Bipolar disorder is a major psychiatric disorder characterized by extreme mood states that alternate between mania and depression. Family, twin, and adoption studies indicate a genetic component to the disease, but the etiology is suspected to be complex, with multiple genes contributing to an increased susceptibility to the disorder. We have previously reported a genome scan in which a genome-wide maximum LOD score indicated evidence of linkage at the marker D22S278 at 22q13. This area is of particular interest since it is also implicated in schizophrenia, and thus may harbor a susceptibility gene common to both disorders. In our further efforts to fine map this region, we examined 10 microsatellite markers spanning an interval of 2.3 MB in a set of 142 parent–proband triads. Linkage disequilibrium to illness was tested using the Transmission Disequilibrium Test. Haplotypes were determined and marker-to-marker linkage disequilibrium across the region was examined. D22S281 and D22S685 yielded suggestive evidence of linkage disequilibrium to bipolar disorder (empirical *P* values of 0.023 and 0.036, respectively), but a marker-to-marker analysis indicates that a higher density screen is needed to adequately analyze this region. Psychiatr Genet 12:231–235 © 2002 Lippincott Williams & Wilkins.

Keywords: bipolar disorder, linkage disequilibrium, human chromosome 22

INTRODUCTION

Bipolar disorder is a severe psychiatric disorder characterized by alternation between extreme mood states. It occurs in approximately 1% of the population, and is associated with psychosis and a substantial risk of suicide (Goodwin and Jamison, 1990). Twin and family studies indicate a genetic component to the disorder, but the non-Mendelian inheritance pattern of the disorder suggests the disease is complex with multiple susceptibility loci (Tsuang and Faraone, 1990). Numerous loci have been implicated by linkage studies but, to date, no specific gene has been definitively identified within these linked regions.

We have recently conducted a genome scan of 20 families with bipolar disorder from the general North American population. These data yielded a genome-wide maximum LOD score of 3.8 at D22S278 (Kelsoe *et al.*, 2001). Surprisingly, this was near the same region implicated in studies of schizophrenia. Subsequently, this overlap of linkage findings in bipolar disorder and schizophrenia has been reported in

several other chromosomal regions and has led to the speculation that these disorders may share some common susceptibility genes (Kelsoe, 1999; Berrettini, 2000).

Based on these results, we have conducted linkage disequilibrium studies to examine the region near D22S278 in greater detail. By fine-mapping the region, we hope to better define the region implicated in bipolar disorder. We now report the results of linkage disequilibrium analysis in 142 triads using the Transmission Disequilibrium Test (TDT) with 10 microsatellite markers in the vicinity of D22S278.

METHODS

Subjects

Families for this study came from two different clinical collections. Thirty-six multiplex families were ascertained through a proband with bipolar I or bipolar II collected at one of three sites (San Diego, Vancouver, and Cincinnati) as part of a collaborative

linkage study of bipolar disorder. All subjects were interviewed using the Structured Clinical Interview for DSM-III-R (Spitzer *et al.*, 1992) for Diagnostic and Statistical Manual of the American Psychiatric Association DSM-III-R or DSM-IV. Diagnoses were determined by consensus review of interview and medical record materials by a panel of experienced clinicians as described previously (Kelsoe *et al.*, 2001). Nuclear families were selected from this sample for linkage disequilibrium studies using the TDT. Thirty-one of these families included a proband and both parents, two included an affected sibpair and both parents, and three families included three affected sibs and both parents.

DNA samples for an additional 106 nuclear families were obtained for subjects selected from multiplex families collected as part of the NIMH Genetics Initiative for Bipolar Disorder. These subjects were ascertained and interviewed using the Diagnostic Interview for Genetic Studies at one of four participating sites (Berrettini *et al.*, 1997; NIMH Genetics Initiative Bipolar Group, 1997). Thirty-eight of these families included a proband and both parents, while 68 included an affected sibpair and both parents.

Genotyping

After obtaining informed consent, blood was drawn on all subjects and immortalized lymphoblastoid cell lines were established. DNA was prepared from cultured cells by phenol/chloroform extraction. Microsatellite markers were selected from the Genome Database (<http://www.gdb.org>). Fluorescently labeled primers were used to amplify markers by polymerase chain reaction. Allele sizes were determined

by electrophoresis on an ABI 377 as described elsewhere (Kelsoe *et al.*, 2001).

Statistical analyses

Linkage studies in complex diseases by nature implicate a large chromosomal region (Terwilliger *et al.*, 1997). However, linkage disequilibrium is subject to association between alleles due to population stratification and admixture rather than from an ancestral association. Family-based tests of linkage disequilibrium such as the TDT are robust to population admixture (Spielman *et al.*, 1993; Ewens and Spielman, 1995).

The extended TDT (ETDT) is a program that performs a TDT analysis for markers with multiple alleles (Sham and Curtis, 1995a). The ETDT employs two alternative approaches to calculate the TDT statistic. The allele-wise analysis examines transmission of alleles across all genotypes. The genotype-wise analysis considers every heterozygous parental genotype separately, and examines whether each allele of the genotype is transmitted to affected offspring. To evaluate the significance of the TDT statistic obtained for each marker, a Monte Carlo approach was employed using 1000 simulations with the MCETDT program (Sham and Curtis, 1995b). Haplotypes were generated using Simwalk2 (Sobel and Lange, 1996), and linkage disequilibrium statistics were calculated with GOLD (Abecasis and Cookson, 2000). Haplotypes could not be definitively determined for two parents.

RESULTS

Using the ETDT program, we analyzed 10 microsatellite markers spanning a 2.3 MB region encompassing

TABLE 1. Extended Transmission Disequilibrium Test results

Marker name	Chromosomal ^a location	Distance (base pairs) ^b	Allele wise	Allele-wise empirical	Genotype wise	Genotype-wise empirical
D22S1162	31106990–31107144	0	0.293	0.354	0.105	0.299
D22S281	31132757–31132897	25 613	0.582	0.634	0.007	0.023
D22S685	31391651–31391837	258 754	0.064	0.078	0.016	0.036
D22S691	31671712–31671955	279 875	0.24	0.324	0.212	0.337
D22S1152	31935156–31935426	263 201	0.9	0.909	0.052	0.074
D22S304	32166729–32166839	231 303	0.227	0.361	0.216	0.41
D22S277	33014905–33015070	848 066	0.84	0.91	0.018	0.129
D22S1142	33017284–33017473	2214	0.943	0.987	0.236	0.509
D22S278	33149958–33150195	132 485	0.19	0.254	0.321	0.53
D22S1173	33379930–33380191	229 735	0.746	0.874	0.903	0.952

^aChromosomal location is based on actual sequence position as reported in the public database at the time of writing, and it is interpolated using downloaded sequence data when actual position is not available (D22S281 and D22S685).

^bDistance is calculated using distance between primers rather than actual distance between repeated elements.

TABLE 2. Marker-to-marker linkage disequilibrium

	Standardized disequilibrium coefficient (D')									
	D22S1162	D22S281	D22S685	D22S691	D22S1152	D22S304	D22S277	D22S1142	D22S278	D22S1173
D22S1162		0.509	0.116	0.133	0.099	0.118	0.148	0.091	0.105	0.089
D22S281	<0.0001		0.142	0.115	0.119	0.088	0.096	0.078	0.057	0.121
D22S685	0.3028	0.0506		0.158	0.14	0.145	0.118	0.101	0.095	0.129
D22S691	0.05	0.6442	0.0016		0.258	0.104	0.142	0.109	0.085	0.103
D22S1152	0.8101	0.4559	0.0651	<0.0001		0.216	0.093	0.069	0.062	0.07
D22S304	0.5038	0.4557	0.0646	0.2525	<0.0001		0.074	0.121	0.076	0.099
D22S277	0.2202	0.6648	0.1334	0.0203	0.8014	0.9245		0.148	0.149	0.125
D22S1142	0.6759	0.8966	0.4471	0.1983	0.7555	0.2864	0.0055		0.278	0.066
D22S278	0.7537	0.9985	0.2535	0.8634	0.5529	0.7817	0.0053	<0.0001		0.111
D22S1173	0.9687	0.3693	0.7928	0.3294	0.3509	0.5826	0.0885	0.8066	0.7489	
	Chi-square P value									

Values above the diagonal are the standardized disequilibrium coefficient (D'). Values below the diagonal are the chi-square P values.

D22S278. As shown in Table 1, suggestive evidence of linkage disequilibrium to bipolar disorder was obtained at D22S281, 2 MB from D22S278 (genotype-wise empirical $P = 0.023$). D22S685, which is located 200 kb telomeric to D22S281, also yielded suggestive evidence of linkage disequilibrium (genotype-wise empirical $P = 0.036$). No other marker examined gave any evidence of linkage disequilibrium.

To interpret these results and to estimate the required marker density for a higher resolution study, we examined marker-to-marker linkage disequilibrium. Table 2 illustrates the standardized disequilibrium coefficient (D') values and the χ^2 values from this analysis. Linkage disequilibrium was not strongly conserved in the 2.3 MB region examined as most of the D' values fall below 0.15. The D' values do not always correspond to P values since a single occurrence of rare alleles for each of the two loci on the same haplotype can produce a significant P value. D' corrects for allele frequencies, thereby preventing an inflation of the D' value resulting from such rare events. Several such instances of the lack of correlation between these measures are apparent in Table 2. As most of our markers were separated by more than 200 kb, these results are not unexpected. However, some close marker pairs did exhibit linkage disequilibrium, although this was not always consistent. In the case of markers D22S1162 and D22S281, significant linkage disequilibrium ($D' = 0.509$) was detected, and these markers are separated by 25 kb. D22S1142 and D22S277 are only separated by 2.2 kb; however, they show much less linkage disequilibrium ($D' = 0.148$).

DISCUSSION

We examined 142 families for linkage disequilibrium in 10 microsatellite markers. Approximately 2 MB centromeric to D22S278, we detected modest evidence of linkage disequilibrium at D22S281 and D22S685. No linkage disequilibrium was detected at marker D22S278. We also examined marker-to-marker linkage disequilibrium in the region, and only modest linkage disequilibrium was found overall. This suggests that a much higher marker density will be required to adequately examine this region for linkage disequilibrium with bipolar disorder.

Many statistical and genetic factors play into examining linkage disequilibrium, and thus these should be considered when interpreting our results. Since the allele-wise test was not significant at any of the markers tested, our results may be more indicative of linkage rather than linkage disequilibrium. Another major limitation to our study is the limited density of markers. Our analysis of marker-to-marker linkage disequilibrium argues that linkage disequilibrium is, in general, not preserved over large distances in this genomic region. Therefore, a much higher marker density than can be provided by microsatellite markers will be required to adequately cover this chromosomal segment and to interpret our linkage disequilibrium results in this region. Furthermore, as microsatellites have a higher mutation rate, it has been argued that linkage disequilibrium may be less likely to be preserved and, therefore, that they are less effective for linkage disequilibrium mapping. This factor, however, must be balanced against their greater informativeness. Finally, allelic heterogeneity

could substantially reduce our power to detect linkage disequilibrium.

Our data show suggestive evidence of linkage disequilibrium at markers D22S281 and D22S685, and although modest, in the context of existing linkage and linkage disequilibrium studies of this region, they provide some support for other reports of a susceptibility locus for bipolar disorder in this region. Detera-Wadleigh *et al.* (1999) reported a maximum LOD score of 2.5 between markers D22S685 and D22S689 in a linkage study of 21 families with bipolar disorder. Mujaheed *et al.* (2000) looked for evidence of linkage disequilibrium between bipolar disorder and D22S278 in an Arab-Palestinian population, and they found modest evidence of linkage disequilibrium ($P = 0.031$). Evidence for linkage with schizophrenia has been found at D22S278 and at nearby markers (Coon *et al.*, 1994; Pulver *et al.*, 1994; Vallada *et al.*, 1995; Gill *et al.*, 1996). Furthermore, D22S278 has also been implicated in linkage disequilibrium studies of schizophrenia (Moises *et al.*, 1995; Schizophrenia Collaborative Linkage Group for Chromosome 22, 1998).

Since this region is implicated in both schizophrenia and bipolar disorder, it may contain a susceptibility gene for both disorders. Only the identification of a disease susceptibility gene and functional variants will enable the definitive testing of this hypothesis. Such a discovery could provide new insight into fundamental pathophysiology and the relationship of the underlying neurobiology of these two disorders.

Acknowledgements

The authors would like to thank the family members who participated in this study, without whom it would not be possible. This work was supported by grants to J.R.K. from the Department of Veterans Affairs and the National Institute of Mental Health (NIMH) (MH47612, MH59567), the UCSD Mental Health Clinical Research Center (MH30914), and the UCSD General Clinical Research Center (M01 RR00827). Support was provided to S.L. by the NIH-sponsored UCSD Genetics Training Program (GM08666). The authors would also like to acknowledge the expert advice and technical assistance of Meghan Alexander, John Brown, Sarah Shaw and Nicholas Schork.

Data and biomaterials were collected in four projects that participated in the NIMH Bipolar Disorder Genetics Initiative. From 1991 to 1998, the Principal Investigators and Co-Investigators were: Indiana University, Indianapolis, IN, USA (UO1 MH46282), John Nurnberger, MD, PhD, Marvin Miller, MD, and Elizabeth Bowman, MD; Washington University, St Louis, MO, USA (UO1 MH46280), Theodore Reich,

MD, Allison Goate, PhD, and John Rice, PhD; Johns Hopkins University, Baltimore, MD, USA (UO1 MH46274), J. Raymond DePaulo, Jr., MD, Sylvia Simpson, MD, MPH, and Cohn Stine, PhD; and NIMH Intramural Research Program, Clinical Neurogenetics Branch, Bethesda, MD, USA, Elliot Gershon, MD, Diane Kazuba, BA, and Elizabeth Maxwell, MSW.

REFERENCES

- Abecasis GR, Cookson WO (2000). GOLD – graphical overview of linkage disequilibrium. *Bioinformatics* **16**:182–183.
- Berrettini WH (2000). Are schizophrenic and bipolar disorders related? A review of family and molecular studies. *Biol Psychiatry* **48**:531–538.
- Berrettini WH, Ferraro TN, Goldin LR, Detera-Wadleigh SD, Choi H, Muniec D, *et al.* (1997). A linkage study of bipolar illness. *Arch Gen Psychiatry* **54**:27–35.
- Coon H, Holik J, Hoff M, Reimherr F, Wender P, Myles-Worsley M, *et al.* (1994). Analysis of chromosome 22 markers in nine schizophrenia pedigrees. *Am J Med Genet* **54**:72–79.
- Detera-Wadleigh SD, Badner JA, Berrettini WH, Yoshikawa T, Goldin LR, Turner G, *et al.* (1999). A high-density genome scan detects evidence for a bipolar-disorder susceptibility locus on 13q32 and other potential loci on 1q32 and 18p11.2. *Proc Natl Acad Sci USA* **96**:5604–5609.
- Ewens WJ, Spielman RS (1995). The transmission/disequilibrium test: history, subdivision, and admixture. *Am J Hum Genet* **57**:455–464.
- Gill M, Vallada H, Collier D, Sham P, Holmans P, Murray R, *et al.* (1996). A combined analysis of D22S278 marker alleles in affected sib-pairs: support for a susceptibility locus for schizophrenia at chromosome 22q12. Schizophrenia Collaborative Linkage Group (Chromosome 22). *Am J Med Genet* **67**:40–45.
- Goodwin FK, Jamison KR (1990). *Manic-Depressive Illness*. New York: Oxford University Press.
- Kelsoe JR (1999). Recent progress in the search for genes for bipolar disorder. *Curr Psychiatry Rep* **1**:135–140.
- Kelsoe JR, Spence MA, Loetscher E, Foguet M, Sadovnick AD, Remick RA, *et al.* (2001). A genome survey indicates a possible susceptibility locus for bipolar disorder on chromosome 22. *Proc Natl Acad Sci USA* **98**:585–590.
- Moises HW, Yang L, Li T, Havsteen B, Fimmers R, Baur MP, *et al.* (1995). Potential linkage disequilibrium between schizophrenia and locus D22S278 on the long arm of chromosome 22. *Am J Med Genet* **60**:465–467.
- Mujaheed M, Corbex M, Lichtenberg P, Levinson DF, Deleuze JF, Mallet J, Ebstein RP (2000). Evidence for linkage by transmission disequilibrium test analysis of a chromosome 22 microsatellite marker D22S278 and bipolar disorder in a Palestinian Arab population. *Am J Med Genet* **96**:836–838.
- NIMH Genetics Initiative Bipolar Group (1997). Genomic survey of bipolar illness in the NIMH genetics initiative pedigrees: a preliminary report. *Am J Med Genet* **74**:227–237.

- Pulver AE, Karayiorgou M, Wolynec PS, Lasseter VK, Kasch L, Nestadt G, *et al.* (1994). Sequential strategy to identify a susceptibility gene for schizophrenia: report of potential linkage on chromosome 22q12-q13.1: Part 1. *Am J Med Genet* **54**:36–43.
- Schizophrenia Collaborative Linkage Group for Chromosome 22 (1998). A transmission disequilibrium and linkage analysis of D22S278 marker alleles in 574 families: further support for a susceptibility locus for schizophrenia at 22q12. *Schizophr Res* **32**:115–121.
- Sham PC, Curtis D (1995a). An extended transmission/disequilibrium test (TDT) for multi-allele marker loci. *Ann Hum Genet* **59**(Part 3):323–336.
- Sham PC, Curtis D (1995b). Monte Carlo tests for associations between disease and alleles at highly polymorphic loci. *Ann Hum Genet* **59**(Part 1):97–105.
- Sobel E, Lange K (1996). Descent graphs in pedigree analysis: applications to haplotyping, location scores, and marker-sharing statistics. *Am J Hum Genet* **58**:1323–1337.
- Spielman RS, McGinnis RE, Ewens WJ (1993). Transmission test for linkage disequilibrium: the insulin gene region and insulin-dependent diabetes mellitus (IDDM). *Am J Hum Genet* **52**:506–516.
- Spitzer RL, Williams JB, Gibbon M, First MB (1992). The Structured Clinical Interview for DSM-III-R (SCID). I: History, rationale, and description. *Arch Gen Psychiatry* **49**:624–629.
- Terwilliger JD, Shannon WD, Lathrop GM, Nolan JP, Goldin LR, Chase GA, Weeks DE (1997). True and false positive peaks in genomewide scans: applications of length-biased sampling to linkage mapping. *Am J Hum Genet* **61**:430–438.
- Tsuang MT, Faraone SV (1990). *The Genetics of Mood Disorders*. Baltimore, MD: Johns Hopkins University Press.
- Vallada H, Curtis D, Sham PC, Murray RM, McGuffin P, Nanko S, *et al.* (1995). Chromosome 22 markers demonstrate transmission disequilibrium with schizophrenia. *Psychiatr Genet* **5**:127–130.