27. R. T. Paine, Nature **355**, 73 (1992).

- 28. O. Sundene, Nytt. Mag. Bot. 9, 155 (1962).
- T. B. Hart, J. A. Hart, P. G. Murphy, Am. Nat. 133, 613 (1989).
- S. D. Torti, P. D. Coley, T. A. Kuser, Am. Nat. 157, 141 (2001).
- 31. R. H. MacArthur, J. W. MacArthur, J. Preer, *Am. Nat.* **96**, 167 (1962).
- 32. D. Tilman, Nature 405, 208 (2000).
- L. Weis and M. O'Connor assisted in data collection; P. Kareiva, D. Schindler, and S. Naeem read early drafts; and J. Ruesink, R. B. Huey, and E. Buhle provided statis-

No Major Schizophrenia Locus Detected on Chromosome 1q in a Large Multicenter Sample

Douglas F. Levinson,^{1*} Peter A. Holmans,² Claudine Laurent,³ Brien Riley,⁴ Ann E. Pulver,⁵ Pablo V. Gejman,⁶
Sibylle G. Schwab,⁷ Nigel M. Williams,⁸ Michael J. Owen,⁸ Dieter B. Wildenauer,⁷ Alan R. Sanders,⁶ Gerald Nestadt,⁵ Bryan J. Mowry,^{9,10} Brandon Wormley,⁴ Stéphanie Bauché,³
Stéphane Soubigou,¹¹ Robert Ribble,⁴ Deborah A. Nertney,⁹ Kung Yee Liang,¹² Laura Martinolich,⁶ Wolfgang Maier,⁷ Nadine Norton,⁸ Hywel Williams,⁸ Margot Albus,¹³
Eric B. Carpenter,⁶ Nicola deMarchi,¹⁴ Kelly R. Ewen-White,¹⁵ Dermot Walsh,¹⁶ Maurice Jay,³ Jean-François Deleuze,¹¹ F. Anthony O'Neill,¹⁷ George Papadimitriou,¹⁸
Ann Weilbaecher,⁶ Bernard Lerer,¹⁹ Michael C. O'Donovan,⁸
Dimitris Dikeos,¹⁸ Jeremy M. Silverman,²⁰ Kenneth S. Kendler,⁴ Jacques Mallet,³ Raymond R. Crowe,²¹ Marilyn Walters²²

Reports of substantial evidence for genetic linkage of schizophrenia to chromosome 1q were evaluated by genotyping 16 DNA markers across 107 centimorgans of this chromosome in a multicenter sample of 779 informative schizophrenia pedigrees. No significant evidence was observed for such linkage, nor for heterogeneity in allele sharing among the eight individual samples. Separate analyses of European-origin families, recessive models of inheritance, and families with larger numbers of affected cases also failed to produce significant evidence for linkage. If schizophrenia susceptibility genes are present on chromosome 1q, their population-wide genetic effects are likely to be small.

Schizophrenia causes severe morbidity in 0.2 to 1% of the world's population, with a heritability of 0.70 to 0.85 attributable to complex inheritance (1). No specific genetic variation has been convincingly associated with susceptibility. Some genome-wide scans have produced significant evidence for linkage, but no result has been consistently replicable (2). In small genome scans of complex disorders, the largest estimated genetic effects often reflect a substantial upward bias, requiring evaluation in independent, larger samples (3). The present multicenter pedigree sample was assembled to determine the degree of support for schizophrenia candidate regions (4, 5).

Several recent reports have suggested schizophrenia susceptibility loci of major effect on chromosome 1q. Brzustowicz *et al.* (6) reported a significant multipoint lod score (logarithm of the odds ratio for linkage) of 6.50 between markers D1S1653 and D1S1679 [162 to 163 cM from the p terminus (7)] in 22 Canadian-Celtic families. Nearby, Gurling *et al.* (8) reported a multipoint lod score of 3.2 (176.6 cM) in 13 British and Icelandic pedigrees. More distally, Ekelund *et al.* (9) reported lod scores of 3.2 (240.4 cM) in 168 Finnish nuclear families, and of 2.30 (222 cM) in 53 families from an isolated subpopulation. Finally, the Disrupted in Schizophrenia (DISC) genes *DISC-1* and *DISC-2* (10) (238.5 cM) are disrupted by a balanced (1;11) (q42.1;q14.3) translocation that segregates with schizophrenia and mood disorders in a Scottish pedigree (11).

To evaluate these findings, we genotyped 16 microsatellite markers (12) on chromosome 1q in 779 informative pedigrees containing 984 affected sibling pairs (ASPs) and 1918 genotyped individuals with schizophrenia or schizoaffective disorder, from eight independently collected samples (13) [Web tables 1 and 2 (14)]. The chromosome 1q findings were reported after formation of the multicenter sample (i.e., there was no selection bias). Primary sta-

tical and other advice. Supported by NSF (grant OCE 9530153) and the Andrew W. Mellon Foundation. I acknowledge with deep gratitude permission from the Makah Indian Nation to work on Tatoosh.

14 January 2002; accepted 8 March 2002

tistical analyses included multipoint ASP (15) and nonparametric linkage (NPL) analyses (16) and logistic regression analysis (17) to test for intersample heterogeneity of sharing in ASPs and for linkage while taking intersample heterogeneity into account (18). Results are shown in Fig. 1 [for details, see Web table 3 (14)]. Only one of the individual samples [National Institute of Mental Health (NIMH)] produced a nominally significant result (P = 0.049) near the Finnish isolate peak (9). We observed no other significant results in individual samples or in the combined sample.

There are several possible explanations for the absence of support for linkage in this large sample, aside from the possibility of undetected genotyping errors or differences in diagnostic practice. Ethnicity could be a factor (19). However, many families in the University of Wales College of Medicine (Cardiff) and Virginia Commonwealth University (VCU)/Ireland samples had ethnic backgrounds (Scottish,

¹Department of Psychiatry, University of Pennsylvania, Philadelphia, PA 19104, USA. ²MRC Biostatistics Unit, Cambridge CB2 2SR, UK. ³Laboratoire de Génétique Moléculaire de la Neurotransmission et des Processus Neurodégénératifs, CNRS, Hôpital de la Pitié Salpêtrière, Paris 75013, France. ⁴Virginia Institute for Psychiatric and Behavioral Genetics, Department of Psychiatry, Virginia Commonwealth University, Richmond, VA 23298, USA. ⁵Department of Psychiatry, Johns Hopkins University School of Medicine, Baltimore, MD 21231, USA. 6 Department of Psychiatry, University of Chicago, Chicago, IL 60637, USA. ⁷Department of Psychiatry, University of Bonn, D-53105 Bonn, Germany. ⁸Department of Psychological Medicine, University of Wales College of Medicine, Cardiff CF14 4XN, UK. 9Queensland Centre for Schizophrenia Research, Wolston Park Hospital, Wacol 4076, Queensland, Australia. ¹⁰Department of Psychiatry, University of Queensland, Brisbane 4029, Queensland, Australia. ¹¹Aventis Pharma SA, Evry 91006, France. ¹²Department of Biostatistics, Johns Hopkins University Bloomberg School of Public Health, Baltimore, MD 21231, USA. ¹³State Mental Hospital, D-85529 Haar, Germany. ¹⁴Clinica Psichiatrica, Seconda Universita degli Studi di Napoli, Naples 80138. Italy. ¹⁵Australian Genome Research Facility. Walter & Eliza Hall Institute of Medical Research, Melbourne, Victoria 3050, Australia. ¹⁶The Health Research Board, Dublin 2, Ireland. ¹⁷Department of Psychiatry, Queens University, Belfast BT7 1NN, Northern Ireland. ¹⁸Department of Psychiatry, University of Athens Medical School, Athens 11526, Greece. ¹⁹Department of Psychiatry, Hadassah–Hebrew University Medical Center, 91120 Jerusalem, Israel. 20Department of Psychiatry. Mt. Sinai School of Medicine. New York, NY 10029, USA. ²¹Mental Health Clinical Research Center and Department of Psychiatry, University of Iowa College of Medicine, Iowa City, IA 52242, USA. ²²Queensland Institute of Medical Research, Herston 4006, Queensland, Australia.

*To whom correspondence should be addressed. Email: dfl@mail.med.upenn.edu Welsh, Irish, Anglo-Saxon) similar to the Celtic Canadian sample. Although the peak Cardiff NPL score (0.92, 160 cM) was near the Canadian peak, any genetic effect would be small; in this sample, the estimate of 53% identical-by-descent (IBD) allele sharing, with 22.7% of ASPs sharing 0 alleles by descent, predicts a locus-specific relative risk to siblings of 1.1. The only nominally significant result observed here was on distal 1q, in the ethnically diverse

REPORTS

NIMH sample. We then analyzed 679 European-origin and 58 African-origin families separately [Web table 4 (14)] (20). For Europeanorigin families, maximum values were $Z_{all} =$ 0.08 (222 cM) and maximum lod score (MLS) = 0.48 (222 cM, 51% IBD sharing). Three samples produced NPL scores of >1.0 [NIMH, 2.35, 222 cM, P = 0.015; Johns Hopkins University (JHU), 1.52, 258.1 cM, P =0.09; Australia/U.S. (AU/US), 1.33, 213.9 cM,



Fig. 1. Results of ASP analysis (**A**) and NPL analysis (**B**). Shown are multipoint MLS (A) and NPL scores (B) for each sample and for all families combined. Applied Biosystems map locations (7) are shown. Locations of reported linkage peaks were extrapolated from markers common to the Marshfield (7) and Applied Biosystems maps—for example, for the Canadian data set (6), the peak at about 168.8 (Marshfield) is shown at 162.5 cM (2 cM proximal to D1S1679); for U.K./Iceland (9) at D1S196 (genotyped here; 176.6 cM); for the Finnish national data set (9) at D1S2709 (240.4 cM); for the Finnish isolate (9) at D1S245 (222.0 cM); and for *DISC-1* (10) near D1S251 [245.05 (Marshfield) shown at 238.52 cM].

P = 0.126); only the NIMH sample produced an MLS value of >1.0 (1.32, 222 cM, 60% IBD sharing, P = 0.032). At the Canadian peak location, no single sample produced an MLS value of >0.16 or an NPL score of >0.91. For African-origin families, maximum values were NPL = 1.25 (193.3 cM, P = 0.18) and MLS = 0.55 (193.3 cM, 56% sharing, P = 0.18). Note that genetic distances among Europeans are small compared with other world populations, although Finns and Icelanders are outliers (21). Our sample does not include comparable families to evaluate the possibility of linkage specifically in Finnish families on distal 1q (9) or in Icelandic families on proximal 1q (8).

There were more affected individuals per family in the Canadian sample (mean = 3.6) than in the present data set (mean = 2.5). Analysis of 209 families with three or more affected cases produced MLS = 0.21 (222.0 cM, 52% IBD sharing) and $Z_{all} = 0.87$ (222.0 cM). The 52 families with four or more cases produced MLS = 0.04 (193.3 cM, 51% IBD sharing) and $Z_{\rm all}$ = 0.46 (258.1 cM). Thus, larger families did not produce evidence for linkage [Web table 5 (14)]. Finally, the MLS in the Canadian study was observed for a parametric analysis under a recessive genetic model. We reanalyzed our data under several recessive genetic models by two-point and multipoint analyses [Web table 6 (14)] (22). The largest heterogeneity lod score (Z_{max}) for the entire sample was 0.31 (multipoint, unaffecteds coded as unknown diagnosis; 185.2 cM), and the largest Z_{max} in an individual sample was 0.65 (multipoint, affecteds only; 210.5 cM, in the VCU/Ireland sample). Our sample would be expected to have 100% power to detect a large genetic effect under the reported recessive model, and to have good power for λ_{sibs} (relative risk to siblings) values of 1.30 or greater (23). Thus, our failure to find evidence for a major schizophrenia susceptibility locus on proximal 1q could not be explained by ethnicity, statistical approach, or pedigree size. The most parsimonious explanation is that the genetic effect reported in the Canadian data set was due to the upward bias caused by maximizing scores across the genome (3), particularly for small data sets and for loci of small effect, because the underlying genetic parameters are being maximized along with the evidence for linkage: "If one assumes that a published locusspecific effect-size estimate ... is accurate ..., one most likely overestimates the power to replicate, perhaps greatly so. . . . A corollary is that failure of replication does not imply that a reported finding is false, even though . . . the locus-specific effect-size estimate from the initial study is likely an overestimate" (3). We cannot determine whether the Canadian finding is a false-positive or a true-positive result whose genetic effect is smaller than reported.

In this large multicenter sample, we were unable to detect a schizophrenia susceptibility locus of major effect on chromosome 1q. It remains possible that the genes identified as disrupted in the Scottish translocation finding (10, 11), or genes in the regions supported by the Finnish (9) and/or Canadian (6) samples, will be shown to have small effects on schizophrenia susceptibility in other populations, or that the pathways in which these genes participate will have more major effects. Identifying such genes to elucidate the pathogenesis of this devastating disorder remains a major goal of schizophrenia research.

References and Notes

- 1. M. T. Tsuang, W. S. Stone, S. V. Faraone, Br. J. Psychiatry 178 (suppl. 40), s18 (2001).
- 2. N. J. Bray, M. J. Owen, Trends Mol. Med. 7, 169 (2001).
- 3. H. H. H. Goring, J. D. Terwilliger, J. Blangero, Am. J.
- Hum. Genet. 69, 1357 (2001).
 Schizophrenia Linkage Collaborative Group, Am. J. Med. Genet. 67, 580 (1996).
- 5. D. F. Levinson *et al.*, Am. J. Hum. Genet. **67**, 652 (2000).
- L. M. Brzustowicz, K. A. Hodgkinson, E. W. C. Chow, W. G. Honer, A. S. Bassett, *Science* 288, 678 (2000).
 Markers and locations are from the Applied Biosys-
- 7. Markers and locations are from the Applied Biosystems (Foster City, CA) high-density map (www. appliedbiosystems.com/products/linkmapping. cfm?prod_id=681&linkmap_id=41). Locations on the Marshfield map (Center for Medical Genetics, Marshfield, WI; http://research.marshfieldclinic.org/ genetics/Map_Markers/maps/IndexMapFrames.html) are typically 5 to 6 cM farther from the p terminus.
- H. M. Gurling et al., Am. J. Hum. Genet. 68, 661 (2001).
- 9. J. Ekelund et al., Hum. Mol. Genet. 10, 1611 (2001).
- 10. J. K. Millar et al., Hum. Mol. Genet. 9, 1415 (2000).
- D. H. Blackwood, Am. J. Hum. Genet. 69, 428 (2001).
 Sixteen fluoresceinated CA-repeat markers covered 107.5 cM of chromosome 1q (sex-averaged genetic distance). Heterozygosity averaged 0.768; intermarker spacing averaged 7.14 cM [see Web table 2 (14) for details]. Primers (Applied Biosystems) were distributed to each laboratory, and optimal conditions were suggested after testing in Cardiff. The AU/US, JHU, and NIMH data sets were genotyped at the Australian Genome Research Facility (Melbourne, Australia).
- 13. The eight samples and references to their methods are as follows: AU/US (24, 25) (molecular methods apply also to JHU and NIMH), University of Bonn (26), Cardiff (27), University of Chicago (28), CNRS (29, 30), JHU (31), NIMH (32) [for a publicly available data set, see (33)], and VCU/Ireland (34, 35). Research diagnostic interviews were completed by research clinicians and best-estimate diagnoses were made based on interviews, records, and informant reports. Affected cases had DSM-IIIR/DSM-IV diagnoses of schizophrenia or schizoaffective disorder. Predominant ethnic origins were as follows: Bonn, German, Israeli/Sephardic; Chicago, AU/US, JHU, and NIMH, European, African American; CNRS, French, French/African/Indian mixtures (Reunion Island): VCU/Ireland, Irish; Cardiff, English, Welsh. The NIMH sample was ethnically diverse. For details of sample sizes, see Web table 1 (14).
- Supplementary material is available on Science Online at www.sciencemag.org/cgi/content/full/296/ 5568/739/DC1.
- 15. P. Holmans, Am. J. Hum. Genet. 52, 362 (1993).
- L. Kruglyak et al., Am. J. Hum. Genet. 58, 1347 (1996).
 D. A. Dorr, J. P. Rice, C. Armstrong, T. Reich, M. Blehar, Genet. Epidemiol. 14, 617 (1997).
- 18. Multipoint ASP and NPL analyses were performed on each data set and on all pedigrees combined (using separate allele frequencies for each data set). Web tables 7 to 9 (14) show linkage scores for all analyses. ASP analyses considered all possible pairs {[5 × (5 - 1)]/2 for

REPORTS

S affected sibs). Region-wide P values were computed empirically by simulating 5000 replicates (assuming no linkage). Logistic regression analyses [Web table 3 (14)] tested intersite heterogeneity in ASP sharing proportions and overall significance of linkage allowing for intersite heterogeneity, with P values based on simulation. See (14) for details. For NPL scores, the $Z_{\rm all}$ scoring function was used (16), which considers allele sharing among all genotyped affected cases in the pedigree including ill siblings, parents, offspring, and other relatives, whereas the MLS statistic considers only sharing within affected sibling pairs.

- 19. J. D. Terwilliger, K. M. Weiss, *Curr. Opin. Biotechnol.* 9, 578 (1998).
- 20. See Web table 4 (14) for detailed results for 679 European-origin and 58 African-origin pedigree subgroups. "Other" ancestries (n = 42; Asian, Micronesian, Indian, Sephardic, or uncertain) were not analyzed separately.
- L. L. Cavalli-Sforza, P. Menozzi, A. Piazza, *The History* and Geography of Human Genes (Princeton Univ. Press, Princeton, NJ, 1994), pp. 268–272.
- 22. Recessive analyses [Web table 6 (14)] used the model associated with the larges Z_{max} for the Canadian sample (6): disease allele frequency = 0.065, f (AA) = 0.50, f (Aa) = f (aa) = 0.0015. GENE-HUNTER 2.0 was used to compute two-point heterogeneity lod scores for D1S484, D1S2878, and D1S196, and multipoint lod scores using 16 markers; multipoint analyses were repeated with disease allele frequency = 0.13 and with unaffected cases coded as diagnosis unknown.
- 23. The recessive model described above (22), assuming 75% of families linked (6), predicts a population-wide λ_{sibs} of 3.55 (36). With 800 ASPs there is 100% power to detect MLS = 3 at $\lambda_{sibs} = 1.8$ (10-cM map), with expected MLS > 20 for $\lambda_{sibs} = 3$ (no parents typed) (37). For comparable families containing 800 ASPs, simulation studies determined power (to detect genome-wide significant linkage) ranging from 66 to 94% (dominant model) as λ_{sibs} varied from 1.27 to 1.36, and from 48 to 68% (recessive) for λ_{sibs} from 1.24 to 1.31 (38).
- 24. D. F. Levinson et al., Am. J. Psychiatry 155, 741 (1998).
- 25. K. R. Ewen et al., Am. J. Hum. Genet. 67, 727 (2000).
- 26. S. G. Schwab et al., Mol. Psychiatry 5, 638 (2000).
- 27. N. M. Williams et al., Hum. Mol. Genet. 8, 1729 (1999).

- 28. Q. Cao et al., Genomics 43, 1 (1997).
- 29. F. Bonnet-Brilhault *et al., Eur. J. Hum. Genet.* **7**, 247 (1999).
- 30. D. Campion et al., Psychiatry Res. 51, 215 (1994).
- 31. J. L. Blouin et al., Nature Genet. 20, 70 (1998).
- C. R. Cloninger et al., Am. J. Med. Genet. 81, 275 (1998).
- 33. Schizophrenia Genetics Initiative Data Archive (http://zork.wustl.edu/nimh/sz.html).
- 34. R. E. Straub *et al.*, *Nature Genet.* **11**, 287 (1995).
- 35. R. E. Straub et al., Am. J. Med. Genet. 81, 296 (1995).
- 36. J. W. James, Ann. Hum. Genet. **35**, 47 (1971).
- 37. E. R. Hauser *et al.*, *Genet. Epidemiol.* **13**, 117 (1996).
- 38. P. A. Holmans, D. F. Levinson, unpublished data.
 - The authors gratefully acknowledge participation of family members as well as the assistance of D. Nancarrow, N. Hayward, D. P. Lennon, M. Gladis, J. Endicott, M. S. O'Brien, C. E. Thornley, and H. L. Jones. Supported by NIMH grant MH61602 (D.F.L., C.L., B.R., A.E.P., P.V.G., D.B.W., M.J.O.). Additional support provided by NIMH grants MH 41953, 52537, and 45390 (B.R. and K.S.K.); the U.K. Medical Research Council (M.J.O.); Deutsche Forschungsgemeinschaft grant SFB 400 (D.B.W., W.M.); the German-Israeli Foundation for Scientific Research (B.L., D.B.W.); NIMH grants KO2-01207 and K24-MH64197 (D.F.L.); National Health and Medical Research Council of Australia (NHMRC) grants 33505 and 35016, Rebecca L. Cooper Medical Research Foundation. Oueensland Department of Health, and NHMRC Network for Brain Research into Mental Disorders (B.J.M.); the NIMH Intramural Program and the Brain Research Foundation, University of Chicago (P.V.G.); NIMH grant RO1-MH57314 (A.E.P.); and CNRS and Aventis Pharma SA (J.M., C.L.). Specimens from the NIMH Schizophrenia Genetics Initiative (NIMH SGI) were used in this study. Data and biomaterials were collected in three projects that participated in the NIMH SGI. From 1991 to 1997, the principal investigators and coinvestigators were Harvard University (grant U01 MH46318) (M. T. Tsuang, S. Faraone, and J. Pepple); Washington University, St. Louis (grant U01 MH46276) (C. R. Cloninger, T. Reich, and D. Svrakic); and Columbia University (grant U01 MH46289) (C. Kaufmann, D. Malaspina, and J. Harkavy Friedman).

16 January 2002; accepted 25 March 2002

Influence of Gene Action Across Different Time Scales on Behavior

Y. Ben-Shahar,¹ A. Robichon,³ M. B. Sokolowski,⁴ G. E. Robinson^{1,2*}

Genes can affect natural behavioral variation in different ways. Allelic variation causes alternative behavioral phenotypes, whereas changes in gene expression can influence the initiation of behavior at different ages. We show that the age-related transition by honey bees from hive work to foraging is associated with an increase in the expression of the *foraging (for)* gene, which encodes a guanosine 3',5'-monophosphate (cGMP)–dependent protein kinase (PKG). cGMP treatment elevated PKG activity and caused foraging behavior. Previous research showed that allelic differences in PKG expression result in two *Drosophila* foraging variants. The same gene can thus exert different types of influence on a behavior.

Some genes influence behavior via genetic polymorphisms, whereas other genes influence behavior via developmental polymorphisms. But little is known about whether the same gene, or orthologs of a gene, can influence behavior in both ways. This knowledge is necessary to develop a comprehensive understanding of how genes and the environment influence behavior, because both involve genomic responsiveness, albeit over vastly different scales of time.

The foraging gene (for) affects naturally